### SHORT COMMUNICATION

# Hot bats: extreme thermal tolerance in a desert heat wave

Artiom Bondarenco · Gerhard Körtner · Fritz Geiser

Received: 7 May 2014 / Revised: 10 June 2014 / Accepted: 17 June 2014 / Published online: 9 July 2014 © Springer-Verlag Berlin Heidelberg 2014

Abstract Climate change is predicted to increase temperature extremes and thus thermal stress on organisms. Animals living in hot deserts are already exposed to high ambient temperatures  $(T_a)$  making them especially vulnerable to further warming. However, little is known about the effect of extreme heat events on small desert mammals, especially tree-roosting microbats that are not strongly protected from environmental temperature fluctuations. During a heat wave with record  $T_{as}$ at Sturt National Park, we quantified the thermal physiology and behaviour of a single free-ranging little broad-nosed (Scotorepens grevii, henceforth Scotorepens) and two inland freetail bats (Mormopterus species 3, henceforth Mormopterus) using temperature telemetry over 3 days. On 11 and 13 January, maximum T<sub>a</sub> was ~45.0 °C, and all monitored bats were thermoconforming. On 12 January 2013, when  $T_a$  exceeded 48.0 °C, Scotorepens abandoned its poorly insulated roost during the daytime, whereas both Mormopterus remained in their better insulated roosts and were mostly thermoconforming. Maximum skin temperatures  $(T_{\rm skin})$  ranged from 44.0 to 44.3 °C in Scotorepens and from 40.0 to 45.8 °C in *Mormopterus*, and these are the highest  $T_{\rm skin}$ values reported for any free-ranging bat. Our study provides the first evidence of extensive heat tolerance in free-ranging desert microbats. It shows that these bats can tolerate the most extreme  $T_{skin}$  range known for mammals (3.3 to 45.8 °C) and delay regulation of  $T_{\rm skin}$  by thermoconforming over a wide temperature range and thus decrease the risks of dehydration and consequently death.

Communicated by: Sven Thatje

A. Bondarenco (⊠) · G. Körtner · F. Geiser Centre for Behavioural and Physiological Ecology, Zoology, University of New England, Armidale NSW 2351, Australia e-mail: bondarenco@gmail.com **Keywords** Arid · Bats · Desert · Extreme heat · Scotorepens · Mormopterus

## Abbreviations

$T_{\rm a}$	Ambient temperature
T <sub>skin</sub>	Skin temperature
T <sub>b</sub>	Body temperature
Scotorepens	Scotorepens greyii (little broad-nosed bat)
Mormopterus	Mormopterus species 3 (inland freetail bat)

#### Introduction

Due to recent increases in global temperature and a predicted rate of further warming at 0.10-0.23 °C per decade (Sánchez-Lugo et al. 2012; IPCC 2013), it is inevitable that more organisms around the world will be exposed to unprecedented high temperature extremes. This is especially challenging for desert organisms which already endure high ambient temperatures  $(T_a)$ . Animals can use several key mechanisms to cope with the increasing global temperature including shifting their current distribution range (Parmesan and Yohe 2003; Pörtner and Farrell 2008), adjusting the timing of diel and/or seasonal activity (Thackeray et al. 2010), acclimating to new environmental conditions during postnatal development (Riek and Geiser 2012) or increasing tolerance to heat stress via natural selection (Feder and Hofmann 1999). However, these mechanisms require a gradual change in  $T_{\rm a}$  to allow animals to adjust their thermal physiology and behaviour to new conditions and therefore do not safeguard against sudden changes in weather and abnormal temperature fluctuations such as cold snaps or heat waves. Such extreme weather anomalies are expected to intensify and become more frequent as a result of current climate change (IPCC 2013). In fact, exposure to short periods of extreme heat can be more dangerous for populations than slowly increasing global temperature (Parmesan et al. 2000) as they can result in mass die-offs and perhaps local extinctions of animals with limited ability to shelter from sudden extreme heat waves such as most birds and many bats (Welbergen et al. 2008; McKechnie and Wolf 2010; Sherwin et al. 2013).

Small, tree-roosting, insectivorous bats appear to be particularly vulnerable to high  $T_{as}$  due to their large surface area to volume ratio, reliance on evaporation for cooling body temperature  $(T_{\rm b})$  without access to drinking water and food during the daytime rest phase and little protection of their roost sites from external  $T_{\rm a}$  and direct solar radiation. Unfortunately, data about physiological and behavioural responses of free-ranging bats to extreme heat (i.e.  $T_a > 40$  °C) are scarce and often limited to visual observations of bats exposed to high temperatures in their roosts without  $T_{\rm h}$  measurements. These field studies have demonstrated that some species of bats, such as Yuma myotis (Myotis yumanensis), Mexican freetail bats (Tadarida brasiliensis), Angolan freetail bats (Mops condylurus) and eastern cave bats (Vespadelus troughtoni) can tolerate roost temperatures that at times exceed 40 °C (Licht and Leitner 1967a; Bronner et al. 1999; Law and Chidel 2007). On the other hand,  $T_{as} > 42$  °C appear to be lethal for flying foxes (Pteropus spp.) that regularly roost in the tree canopy directly exposed to temperature extremes and sun (Welbergen et al. 2008).

Here, we report the effect of extreme high  $T_a$  on thermal physiology and behaviour of one adult free-ranging little broad-nosed bat (*Scotorepens greyii*, Vespertilionidae, henceforth *Scotorepens*) and two adult inland freetail bats (*Mormopterus* species 3 (Adams et al. 1988), Molossidae, henceforth *Mormopterus*) during the most extreme hot weather in the past four decades in the Australian arid zone (Bureau of Meteorology 2013). These bats had been tagged during the heat wave and therefore provided a unique opportunity to examine their thermal biology and behaviour. While *Scotorepens* is found throughout arid and semi-arid regions and also subtropics and tropics of northern Australia, the distribution of *Mormopterus* is restricted to desert habitats (Churchill 2008).

#### Methods

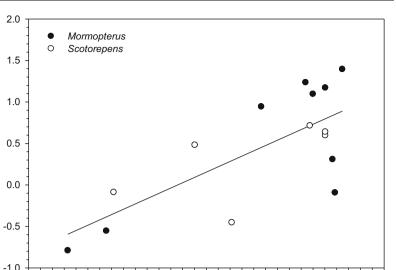
Measurements were made at Sturt National Park near Mt Wood Homestead (29° 28' S, 142° 14' E; ~22 km east from Tibooburra; elevation ~183 m) located in the north-western corner of New South Wales (NSW), Australia, during 11–13 January 2013 (austral summer). The study area contains sparse, open woodlands along creek beds dominated by river red gum (*Eucalyptus camaldulensis*), coolabah (*Eucalyptus coolabah*) and gidgee (*Acacia cambagei*). During the tracking period, drinking water was available from two dams that capture flood water from a creek.

The climate of Sturt National Park is arid, and the longterm average daily  $T_a$  (93–103 years) during the hottest month (January) ranges from 22.1 to 36.2 °C. Temperatures in the shade above 45 °C are rare at the research site and have only been recorded on 66 days (~0.17 %) over 104 years (Tibooburra Post Office, Bureau of Meteorology, Australia). The Australian summer 2012–2013 (December–February) was the hottest on record since 1972–1973 with a prolonged hot spell observed in the country from 25 December to 18 January (Bureau of Meteorology 2013). Furthermore, January 2013 was the ninth warmest January since 1880 in the world (NOAA 2013).

Bats (one female *Scotorepens*, capture body mass=7.2 g; one male and one female *Mormopterus*, body mass= $7.8\pm$ 0.5 g) were captured between 9-11 January 2013, without knowledge of the imminent extreme heat, and fitted with external temperature-sensitive radio transmitters (0.43 g, LB-2NT, Holohil Systems Inc., Carp, Ontario, Canada) following Bondarenco et al. (2013). Before attachment, transmitters were calibrated in a water bath against a precision mercury thermometer (±0.1 °C) traceable to a national standard over a temperature range of 10–42 °C. Skin temperature ( $T_{skin}$ ) was then calculated using third-order polynomial regressions (all  $R^2$ =0.99) with all measured values falling within 0.5 °C of the calibration curve. For small animals, such as bats, resting  $T_{\rm skin}$ measured with external transmitters is usually within 2-3 °C of core  $T_{\rm b}$ , with  $T_{\rm skin}-T_{\rm b}$  differential decreasing with an increase in  $T_a$  (Audet and Thomas 1996; Barclay et al. 1996). To establish whether this is also correct at  $T_a$ s>40 °C, we also measured  $T_{\rm skin}-T_{\rm b}$  differential of six *Scotorepens* and nine Mormopterus under laboratory conditions at T<sub>a</sub>s 40-42 °C using calibrated external radio transmitters (LB-2NT) attached as described above and a calibrated thermocouple thermometer (HH-71T, Omega Engineering, Stamford CT, USA) used to measure core  $T_{\rm b}$  rectally to the nearest 0.1 °C. We found that  $T_{\rm skin}-T_{\rm b}$  differential was similar for both species, and it was only 0.4±0.7 °C at  $T_a$  41.4±0.8 °C ( $T_{skin}$  41.3±1.2 °C). The  $T_{\rm skin}-T_{\rm b}$  differential did not differ between the two species (intercept p < 0.985) and increased steadily with the rise in  $T_a$ with the maximum error of only 1.4 °C at  $T_a$  42.1 °C (Fig. 1). Therefore, we are confident that  $T_{skin}$  measured in the field with external transmitters provides a good representation of  $T_{\rm b}$ . It should, however, be acknowledged that potentially T- $_{skin}-T_b$  differential could be higher at  $T_as >42$  °C due to ambient heat affecting externally attached transmitters.

Individual bats were radiotracked daily to their roosts shortly after sunrise. After locating the roost tree, we used a coaxial cable attached to a 4-m long fishing rod connected to a receiver (Icom, IC-R10, Osaka, Japan) to find the exact bat location. For each animal, a receiver/data logger was placed near the roost tree to remotely record  $T_{\rm skin}$  data every 10 min

Fig. 1 The  $T_{skin}-T_b$  differential measured under laboratory conditions for nine *Mormopterus* and six *Scotorepens* plotted against  $T_a$  (linear mixed effects model— $t_{11}$ =3.14, p=0.009,  $R^2$ = 0.99; n=15, N=15). The intercept was indistinguishable for both species (p<0.985)



41.0

 $T_a (°C)$ 

41.5

42.0

(Körtner and Geiser 1998). During and after the heat wave, we also searched for dead bats at the Mt Wood Homestead complex and Tibooburra with the help of NPWS staff.

T<sub>skin</sub>-T<sub>b</sub> differential

39.5

40.0

40.5

 $T_{\rm a}$  was measured at 10-min intervals with temperature/ humidity data loggers (resolution 0.0625 °C; iButton Hygrochron, DS1923, Maxim Integrated Products Inc., Sunnyvale, California, USA) placed at two locations in the shade 2 m above the ground. Both data loggers provided similar  $T_{\rm a}$ s, and only  $T_{\rm a}$  from a data logger located within 300 m of occupied roosts was used in this study. Prior to use, iButtons were calibrated following Bondarenco et al. (2013).

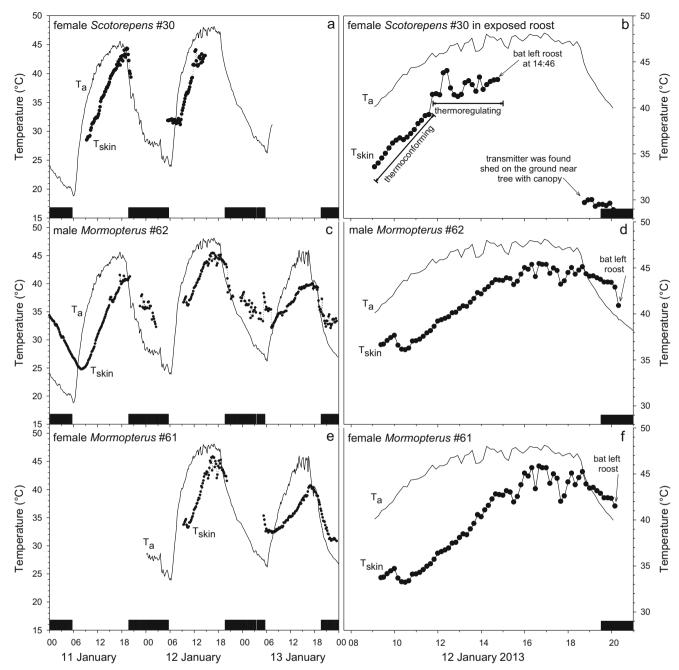
The linear mixed effects model with *Mormopterus* as a reference group and species as a factor was computed to analyse differences in the  $T_{skin}-T_b$  differential between two species with individual bats entered as a random effect to account for repeated measurements (Fig. 1). General linear modelling was used to compare a heating rate among bats with  $T_{skin}$  as a dependent variable, time as an independent variable and individuals as a factor. All models were computed in R (R development Core Team 2012; package "nlme"), and the residuals of the models were checked for homoscedasticity and normality by analysing residual plots. We report numerical values as means  $\pm$  SD, "*n*" for the number of individuals and "*N*" for the number of measurements.

## Results

On the hottest day, 12 January, maximum  $T_a$  reached 48.1 °C, and mean daily  $T_a$  was 37.7 °C. Temperature conditions on 11 and 13 January were similar and only marginally cooler; maximum  $T_a$  was 45.6 and 45.9 °C, and mean daily  $T_a$  was 33.4 and 34.9 °C, respectively. 42.5

On 12 January, Scotorepens roosted inside a slender branch (circumference ~30 cm), ~2 m above the ground, which was exposed to direct sunlight. On this day (Fig. 2a, b), T<sub>skin</sub> passively followed  $T_a$  with a ~2-h delay from  $T_{skin}$  31.8 °C in the morning until  $T_{\rm skin}$  reached the maximum 44.0 °C ( $T_{\rm a}$  46.1 °C). We define this lack of regulatory response to the increasing  $T_{\rm a}$  as thermoconforming behaviour as opposite to a regulated  $T_{\rm skin}$ (see below; Fig. 2b). After T<sub>skin</sub> of Scotorepens reached its maximum, even though  $T_a$  continued to increase,  $T_{skin}$  began to decrease and to fluctuate (i.e. T<sub>skin</sub> was regulated), presumably, indicating active thermoregulation leading to elevated heat dissipation (Fig. 2b). During this time, *Scotorepens* cooled its  $T_{skin}$ from 44.0 °C ( $T_a$  46.1 °C) to 41.2 °C ( $T_a$  47.0 °C), and at the maximum T<sub>a</sub> 48.0 °C, T<sub>skin</sub> was 42.5 °C (5.5 °C below T<sub>a</sub>). Nevertheless, Scotorepens abandoned its roost 3.2 h after the start of active thermoregulation, at 14:46 ( $T_{skin}$  43.0 °C;  $T_a$ 47.3 °C). The transmitter was later found on the ground  $\sim 100$  m from the original roost under an alive tree with exfoliating bark and a well-developed tree canopy providing shade where, presumably, this bat was sheltering.

On the same day (12 January), the male *Mormopterus* was roosting in the partly alive main trunk of a river red gum (circumference ~190 cm), and the female was found in a smaller dead branch (circumference ~110 cm) adjacent to the main trunk of the same tree. Both roosts were ~6 m above the ground and shaded by branches and leaves. On this day,  $T_{\rm skin}$  patterns of both individuals were similar, but the amplitude of  $T_{\rm skin}$  fluctuations differed between the two individuals (Fig. 2c–f).  $T_{\rm skin}$  of the female roosting in the branch increased from a minimum of 33.2 °C in the morning to a maximum of 45.8 °C ( $T_{\rm a}$  47.9 °C);  $T_{\rm skin}$  of the male found in the tree trunk increased from a minimum of 36.1 °C to a maximum of 45.4 °C ( $T_{\rm a}$  47.6 °C).



**Fig. 2** Fluctuations in  $T_{skin}$  (lower trace, dotted line) and  $T_a$  (upper trace, solid line) of the female Scotorepens (a), male (c) and female (e) Mormopterus during the heat wave on 11–13 January 2013; detailed

 $T_{\rm skin}/T_{\rm a}$  recordings for the female *Scotorepens* (**b**), male (**d**) and female (**f**) *Mormopterus* during the extreme hot weather on 12 January 2013. *Black bars* represent night

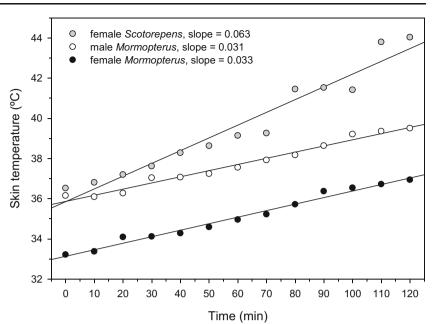
On the previous day, 11 January, the female Scotorepens

was roosting inside a sun-exposed old fence post (circumfer-

ence ~100 cm), ~1 m above the ground. During much of this

Interestingly,  $T_{skin}$  of both *Mormopterus* reached its maximum 4.2 h later (16:40) than in *Scotorepens* (12:26). Hence,  $T_{skin}$  was tracking  $T_a$  with a ~5-h delay (Fig. 2d, f). Furthermore, the mean rate of  $T_{skin}$  increase (heating rate) in *Scotorepens* was about double (0.063 °C min<sup>-1</sup>; p<0.001) and significantly higher (general linear model— $F_{5.33}$ =331.2, p<0.001,  $R^2$ =0.98; n=3, N=39) than in the male (0.031 °C min<sup>-1</sup>; p<0.001) and female (0.033 °C min<sup>-1</sup>; p<0.001) *Mormopterus* (Fig. 3).

Fig. 3  $T_{skin}$  of the female Scotorepens, male and female Mormopterus over 120 min calculated from the time  $T_{skin}$  of all three individuals was steadily increasing (~10:30) until  $T_{skin}$  of Scotorepens reached its maximum and started to decrease (12:26) during the extremely hot 12 January 2013. Note the steeper slope of this relationship in Scotorepens than Mormopterus indicating poor insulation of Scotorepens' roost



whereas on the hottest day,  $T_a$  was still increasing when  $T_{skin}$  maximum was reached. During the same day, the male *Mormopterus* was roosting in a trunk of a dead coolabah (circumference ~130 cm), ~3 m above the ground. The bat was torpid with a minimum  $T_{skin}$  of 24.8 °C in the morning, and it was thermoconforming for most of the day with  $T_{skin}$  tracking  $T_a$  with a ~5.5-h delay (Fig. 2c); maximum  $T_{skin}$  was 41.5 °C ( $T_a$  45.6 °C).

On 13 January, both *Mormopterus* occupied the same roosts as on 12 January. Bats were thermoconforming throughout this day, and the maximum  $T_{skin}$  was 40.7 °C ( $T_a$  43.6 °C) in the female and 40.0 °C ( $T_a$  40.7 °C) in the male *Mormopterus*.  $T_{skin}$  of the female *Mormopterus* was tracking  $T_a$  with a ~5.5-h delay, and  $T_{skin}$  of the male *Mormopterus* was lagging ~6.0 h behind  $T_a$  (Fig. 2c, e).

Although the tagged *Scotorepens* likely and *Mormopterus* definitely survived the extreme heat wave, one recently dead little pied bat (*Chalinolobus picatus*; body mass ~6 g; arid to subtropical distribution range) was found on the floor in a house in Tibooburra on 11 January. Another recently dead animal, Gould's wattled bat (*C. gouldii*; body mass ~10 g; distribution wide range), was found hanging on the mesh of a floor vent at Mt Wood Homestead a few days after the heat wave on 23 January. Several other non-identified dead microbats (likely *C. picatus* based on the description from NPWS staff) were also found during the heat wave inside buildings in Tibooburra.

#### Discussion

Our study provides the first insight into the physiological response of three individuals of small free-ranging arid zone

bats to an extreme heat event. Our data show that appropriate roost selection can be a key factor for the survival of bats during severe heat. Nevertheless, both Scotorepens and *Mormopterus* had to endure a substantially elevated  $T_{skin}$ and  $T_{\rm b}$ . The  $T_{\rm skin}$  values observed in both species were the highest reported for any free-ranging bat and, considering the differential between  $T_{\rm skin}$  and  $T_{\rm b}$  of ~0.4 °C, were above the measured lethal  $T_{\rm b}$  for most placental mammals (42–44 °C; Schmidt-Nielsen 1997). It is also astonishing that Mormopterus can tolerate enormous seasonal Tskin fluctuations ranging from a minimum of only 3.3 °C in winter (Bondarenco 2014) to a maximum of 45.8 °C reported here, giving it a total  $T_{skin}$  range of 42.5 °C. To our knowledge, this is the most extreme  $T_{skin}$  range observed in any mammal and, taking into account  $T_{\rm skin}-T_{\rm b}$  differential, appears to exceed even the widest known  $T_{\rm b}$  fluctuations among endotherms reported for arctic ground squirrels (Spermophilus parryii, total  $T_{\rm b}$  range ~41 °C; Barnes 1989).

While quantitative data on  $T_{\rm b}$  of bats exposed to extreme heat measured under natural conditions are not available, visual observations suggest that heat tolerance does differ among species. For example, in a mixed species roost in a barn loft, pallid bats (*Antrozous pallidus*) moved away from the ceiling areas when  $T_{\rm a}$  reached only 38 °C, whereas Yuma myotis (*Myotis yumanensis*) did not move until  $T_{\rm a}$  at the ceiling beams exceeded 40 °C. Finally, Mexican freetail bats (*Tadarida brasiliensis*) only retreated to the cooler parts of the roost after  $T_{\rm a}$  of beams reached 41–42 °C (Licht and Leitner 1967a).

In comparison, although we could not observe bat activity inside roosts,  $T_{skin}$  data show that *Scotorepens* and *Mormopterus* were mostly thermoconforming during the hot days on 11 and 13 January. If thermoregulation, evaporative cooling or movement did occur during this time,  $T_{skin}$  patterns were not obviously affected. Another approach, employed by bats studied here, was using torpor (see also Bondarenco et al. 2013) or at least a substantial reduction in  $T_b$  in the early morning. Reduced  $T_{skin}$  during thermoconforming behaviour and torpor is usually associated with low metabolic heat production and water loss. This is especially crucial for the survival of small endotherms as less heat needs to be dissipated and, therefore, hyperthermia is delayed or prevented, which will save water that may be required for evaporative cooling during the hottest part of the day (Schmidt-Nielsen et al. 1957; Licht and Leitner 1967b; Bradley and Yousef 1972).

In contrast, when exposed to the extreme heat on 12 January, Scotorepens only initially thermoconformed, but when  $T_{\rm skin}$  reached ~44.0 °C, the bat appeared to commence active thermoregulation, possibly in conjunction with a retreat to a cooler part of the roost. Such increased thermoregulatory effort has to involve active evaporative cooling such as panting, saliva spreading and/or wing flapping (Reeder and Cowles 1951; Licht and Leitner 1967b; Withers 1992; Schmidt-Nielsen 1997; Speakman and Thomas 2003). Heating for both Mormopterus commenced later and at the slower rate. Overall, it appears that both Mormopterus were mostly thermoconforming even during 12 January. Although eventually T<sub>skin</sub> in Mormopterus also started to fluctuate, the fluctuation patterns were similar for bats and were probably caused by variations in  $T_a$ /sun exposure, due to changes in wind, cloud cover or by canopy shading rather than commencement of active T<sub>skin</sub> regulation. Evidently, the largediameter tree trunks and branches used by Mormopterus provide better insulation against extreme temperatures than slender ones, and alive trees provide more insulation and even cooling than dead cracked ones due to differences in water content/flow (Kunz and Lumsden 2003; Briscoe et al. 2014), thus resulting in delayed and slower heating. Moreover, the presence of a developed tree canopy and branches provide additional protection from direct sunlight. Therefore,  $T_{\rm skin}$  of Scotorepens, roosting in a smaller dead coolabah without canopy, was increasing earlier and at a higher rate, forcing this bat to commence heat dissipation earlier and eventually to abandon its roost to avoid pathological hyperthermia when environmental conditions became extreme. For a bat, leaving a roost in daylight, especially during hot weather, is associated with extreme risks firstly by overheating during flight (Voigt and Lewanzik 2011) and secondly by predation from birds (Speakman et al. 1994). Obviously, such behaviour was the last option available to the Scotorepens. Moreover, dead bats found inside buildings suggest that even man-made structures do not always provide enough insulation from extreme heat.

While appropriate roost selection appears to be a key factor for bats' survival under severe heat stress conditions, for those roosting in trees and ceilings, hyperthermia cannot be avoided altogether. The ability of tree-roosting desert bats to tolerate moderate hyperthermia will delay or prevent the onset of evaporative cooling, which is important as these bats have no access to drinking water or food during the daytime and have to rely entirely on the body fluids for heat dissipation. Nevertheless, the provision of artificial watering points at our study site and throughout the Australian arid zone provides desert bats with an opportunity to restore water balance quickly at dusk. Similarly to other water-dependent species across arid and semi-arid Australia (James et al. 1999), this may be especially important for the survival of those bat species that are not desert specialists, such as *Scotorepens*, which is usually found near water (Churchill 2008).

In conclusion, our data show that in contrast to many fruit bats that often roost in the open, microbats can have a better chance of overcoming extreme heat events because of roost protection against radiation, the ability to tolerate high  $T_b$  and opportunistic use of torpor or at least a low  $T_b$  in the morning (Geiser et al. 2011; Bondarenco et al. 2013). However, despite these adaptations, even desert tree-roosting bats can only deal with a limited increase in  $T_a$  as demonstrated by the dead bats found. This shows that small, tree-roosting microbats are not immune from the negative effects of climate change and, because they already experience  $T_b$ s near their thermal limit, may be among the first mammalian species that will be adversely affected by extreme heat events in arid areas around the world.

**Acknowledgments** We thank the National Parks and Wildlife Service (NPWS) staff especially Ingrid Witte, Barb Hawerkamp and Dan Hough for their help with organising the field work and Kim Piddington for the assistance with collecting dead bats. The research was conducted under permits from the NPWS and the UNE Animal Ethics Committee. Financial support was received from the University of New England to AB and the Australian Research Council to FG. Accommodation and transport at Sturt National Park were provided by the NPWS.

#### References

- Adams M, Reardon TR, Baverstock PR, Watts CHS (1988) Electrophoretic resolution of species boundaries in Australian Microchiroptera. IV. The Molossidae (Chiroptera). Aust J Biol Sci 41:315–326
- Audet D, Thomas DW (1996) Evaluation of the accuracy of body temperature measurement using external radio transmitters. Can J Zool 74:1778–1781
- Barclay RMR, Kalcounis MC, Crampton LH, Stefan C, Vonhof MJ, Wilkinson L, Brigham RM (1996) Can external radiotransmitters be used to assess body temperature and torpor in bats? J Mammal 77:1102–1106
- Barnes BM (1989) Freeze avoidance in a mammal: body temperatures below 0 °C in an arctic hibernator. Science 244:1593–1595
- Bondarenco A (2014) Torpor and thermal energetics in Australian arid zone bats. University of New England, Armidale. PhD thesis.

- 183:1113–1122
  Bradley WG, Yousef MK (1972) Small mammals in the desert. In: Yousef MK, Horvath SM, Bullard RW (eds) Physiological adaptations: desert and mountain. Academic, New York, pp 127–142
- Briscoe NJ, Handasyde KA, Griffiths SR, Porter WP, Krockenberger A, Kearney MR (2014) Tree-hugging koalas demonstrate a novel thermoregulatory mechanism for arboreal mammals. Biol Letters. doi: 10.1098/rsbl.2014.0235
- Bronner GN, Maloney SK, Buffenstein R (1999) Survival tactics within thermally-challenging roosts: heat tolerance and cold sensitivity in the Angolan free-tailed bat, *Mops condylurus*. S Afr J Zool 34:1–10
- Bureau of Meteorology (2013). Special climate statement 43—extreme heat in January 2013.
- Churchill S (2008) Australian bats, 2nd edn. Allen & Unwin, Crows Nest
- Feder ME, Hofmann GE (1999) Heat-shock proteins, molecular chaperones, and the stress response: evolutionary and ecological physiology. Annu Rev Physiol 61:243–282
- Geiser F, Stawski C, Bondarenco A, Pavey CR (2011) Torpor and activity in a free-ranging tropical bat: implications for the distribution and conservation of mammals? Naturwissenschaften 98:447–452
- IPCC (2013) Climate change 2013: the physical science basis. Working group I contribution to the fifth assessment report of the Intergovernmental Panel on Climate Change. Summary for policymakers. IPCC
- James CD, Landsberg J, Morton SR (1999) Provision of watering points in the Australian arid zone: a review of effects on biota. J Arid Environ 41:87–121
- Körtner G, Geiser F (1998) Ecology of natural hibernation in the marsupial mountain pygmy-possum (*Burramys parvus*). Oecologia 113: 170–178
- Kunz TH, Lumsden LF (2003) Ecology of cavity and foliage roosting bats. In: Kunz T, Fenton B (eds) Bat ecology. University of Chicago Press, Chicago, pp 3–89
- Law BS, Chidel M (2007) Bats under a hot tin roof: comparing the microclimate of eastern cave bat (*Vespadelus troughtoni*) roosts in a shed and cave overhangs. Aust J Zool 55:49–55
- Licht P, Leitner P (1967a) Behavioral responses to high temperatures in three species of California bats. J Mammal 48:52–61
- Licht P, Leitner P (1967b) Physiological responses to high environmental temperatures in three species of microchiropteran bats. Comp Biochem Physiol 22:371–387
- McKechnie AE, Wolf BO (2010) Climate change increases the likelihood of catastrophic avian mortality events during extreme heat waves. Biol Letters 6:253–256

- NOAA (2013) Global summary information—January 2013. National Oceanic and Atmospheric Administration. http://www.ncdc.noaa. gov/sotc/global/2013/1. Accessed 04 February 2014
- Parmesan C, Root TL, Willig MR (2000) Impacts of extreme weather and climate on terrestrial biota. Bull Am Meteorol Soc 81:443–450
- Parmesan C, Yohe G (2003) A globally coherent fingerprint of climate change impacts across natural systems. Nature 421:37–42
- Pörtner HO, Farrell AP (2008) Physiology and climate change. Science 322:690–692
- R development Core Team (2012) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Reeder WG, Cowles RB (1951) Aspects of thermoregulation in bats. J Mammal 32:389–403
- Riek A, Geiser F (2012) Developmental phenotypic plasticity in a marsupial. J Exp Biol 215:1552–1558
- Sánchez-Lugo A, Kennedy J, Berrisford P (2012) Surface temperature. State of the climate in 2011. Bull Am Meteorol Soc 93:S14–S15
- Schmidt-Nielsen K (1997) Animal physiology. Cambridge University Press, Cambridge
- Schmidt-Nielsen K, Schmidt-Nielsen B, Jarnum SA, Houpt TR (1957) Body temperature of the camel and its relation to water economy. Am J Physiol 188:103–112
- Sherwin HA, Montgomery WI, Lundy MG (2013) The impact and implications of climate change for bats. Mammal Rev 43:171–182
- Speakman JR, Thomas DW (2003) Physiological ecology and energetics of bats. In: Kunz TH, Fenton MB (eds) Bat ecology. University of Chicago Press, Chicago, pp 430–490
- Speakman JR, Lumsden LF, Hays GC (1994) Predation rates on bats released to fly during daylight in south-eastern Australia. J Zool 233: 318–321
- Thackeray SJ, Sparks TH, Frederiksen M, Burthe S, Bacon PJ, Bell JR, Botham MS, Brereton TM, Bright PW, Carvalho L, Clutton-Brock T, Dawson A, Edwards M, Elliott JM, Harrington R, Johns D, Jones ID, Jones JT, Leech DI, Roy DB, Scott WA, Smith M, Smithers RJ, Winfield IJ, Wanless S (2010) Trophic level asynchrony in rates of phenological change for marine, freshwater and terrestrial environments. Glob Change Biol 16:3304–3313
- Voigt CC, Lewanzik D (2011) Trapped in the darkness of the night: thermal and energetic constraints of daylight flight in bats. Proc R Soc B Biol Sci 278:2311–2317
- Welbergen JA, Klose SM, Markus N, Eby P (2008) Climate change and the effects of temperature extremes on Australian flying-foxes. Proc R Soc B 275:419–425
- Withers PC (1992) Comparative animal physiology. Saunders College Publishing, Philadelphia, Pennsylvania